

**NORTHRUP EXHIBIT P**

Crit  
Results

M. Allamay

#7

Notes ( Signal book of several (not all) results photos with this pen (other (on front) was non permanent ink)

- PCR (HTV - MSP) worked well in integrated-heater device, gel electrophoresis verified product. Some, but minimal power (esp. due to known fact that device react in mixture cycled 1-2 times, then at R.T. for  $\frac{1}{2}$  hr & power to 20 cycles due to need to re-solubl corrections - new rxn mixture (total) was added)
- was able to extract  $\sim 100\%$  of aqueous phase with 200  $\mu$ l (set at 30  $\mu$ l) pipette & load 5-6 wells of electrophoresis channel
- (X)  $\rightarrow$  Calculate power consumed in today's experiment compare to batteries

Other Discussion

Last Tues w/ Gary Manilla here (Cetus) along w/  
Russ Higuchi, Bob Watson, Russ's  
technician, myself we tried  
homogeneous detection w/ video  
CCD over 460 thermal cycles

- pulsed Ne - laser (ILEE laser company, Switz) was tried
  - $\rightarrow$  See LLNL Book (notebook) for details

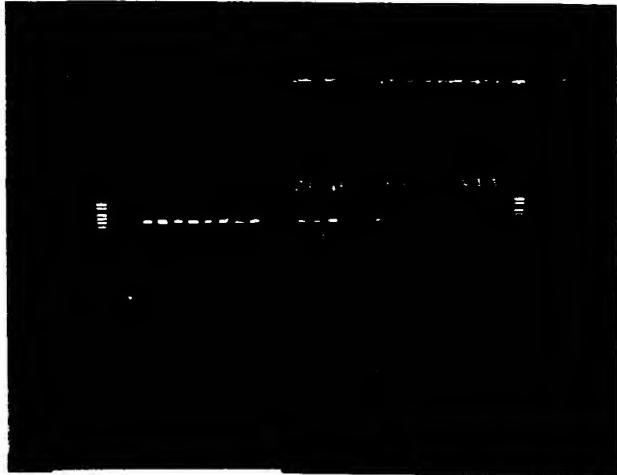
Conf

Results (photos)

Devia PCP results positive

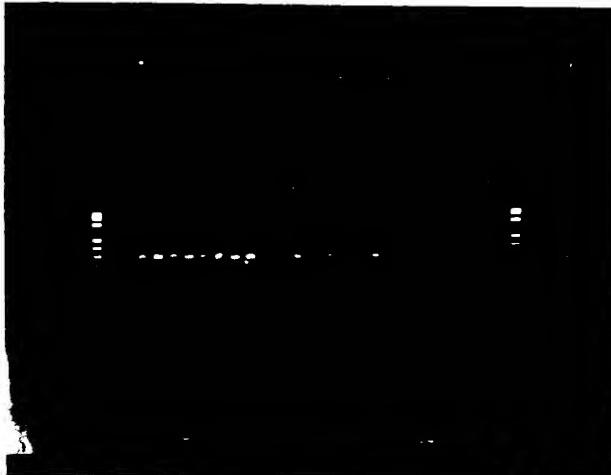
M. All <sup>15</sup>/Aug

elctr. T = 15 min



M. All <sup>15</sup>/Aug T=2 sec  
4.6 > 200

elctr = 40 min



M. All <sup>15</sup>/Aug T=2 sec  
4.6 > 200

taped recipe from  
Watson

K

Rxn	50 $\mu$ l 10X RM	2
par		3
B. Watson	50 $\mu$ l 1 mM dATP	
	50 $\mu$ l M13	4
	(10 $\mu$ l 10X = 100 patches) $\mu$ l/patch	5
M. Alln/long	10 $\mu$ l patch	6
	2.5 $\mu$ l (12.5 $\mu$ l/patch 12.5 12.5/ $\mu$ l/patch = 2 + GAG)	
	327.5 $\mu$ l total	7
	-----	
	500	8

$\theta$  Cetus M. Allfrey

Tiny new PCR system  
(more Temp forgiving)

142 bp product target as ss m13 from  
gag-region of HIV

1) Starting target =  $10^8$  copies in 5  $\mu$ l

$T = 96 - 55 \downarrow 16 - 18$  cycles  
(works at 88+)  
is plenty

2) primers

<u>old names:</u>	<u>new names</u>	
SK 145	= ph 07	$10 \mu\text{M}/\mu\text{l}$
SK 431	= ph 08	

Reaction mixture: (500  $\mu$ l)

50  $\mu$ l 10x Buffer w/ mgel

" 1 mM dNTPs

" m13 w/ gag region of HIV

10  $\mu$ l =  $10 \times 10 = 100$  pmoles

10  $\mu$ l (same Fwd) ph 07 ?

$2\frac{1}{2} \mu\text{l} = 10 \times 1.25 \mu\text{M}/\mu\text{l}$  12.5

Tag  
327.5  $\mu\text{l H}_2\text{O}$

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500  $\mu$ l total run volume

500.0  
172.5  
327.5

M. Alm / *of*

(cont.)

- i) re-use voltage (same device) as  
on March 30 [ie 3.17 V + 98°C]  
at 0.2A

Do only 20 cycles

A) Standards

10, 10, 20, 20, 30, 30, 40, 40

$\sim 1.5 \mu\text{l}$  oil (1-8)

B) Device 30  $\mu\text{l}$  w  $\sim 90 \mu\text{l}$  oil

1-minute cycles at 3.17V  
20 - 1 minute cycles (A-F) 0.2A

electrophoresis

well-problem

run

(f)

P std 6.6 1 2 3 4 5 6 7 8 ++ ABC DDE + 9 10

- 1c) Had to re-solder device  $\checkmark$  after  $\frac{1}{2}$ -cycles  
fix time  $\approx \frac{1}{2}$  hour when was  
at room temp

wire connectors

Results - (1) formed product in both

stds and in wells

(2) wells (and 1) std had

less bright primer-dimers

(3) dev. contained  $\sim 6-5 \mu\text{l}$  gel

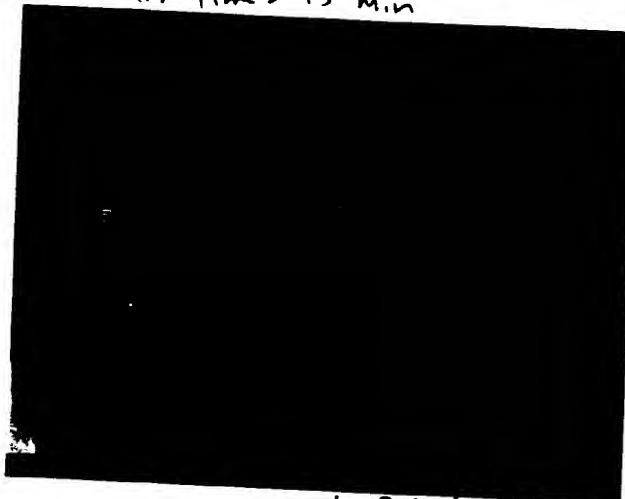
See next  
2 pages:

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Cont results (photos)

M. Ollie

electr. Time = 15 min



T = 1 sec 4.6 3200



T = 1 sec 5.6 3200

#1 Loaned to Milk Ching